

# 2x qPCRBIO Probe Mix Lo-ROX

## Product description:

Combined with the latest advancements in polymerase technology and advanced buffer chemistry qPCRBIO Probe Mix offers market leading performance with minimal optimisation.

PCR Biosystems real-time PCR probe mixes have been designed for use on a wide range of probe technologies including TaqMan®, Scorpions® and molecular beacon probes.

qPCRBIO Probe Mix uses antibody-mediated hot start technology that prevents the formation of primer-dimers to improve reaction sensitivity and specificity.

High-throughput screening has resulted in a buffer system that allows efficient amplification from GC-rich and AT-rich templates, under fast and standard cycling conditions.

2x qPCRBIO Probe Mix Lo-ROX
1 x 1ml
5 x 1ml
20 x 1ml
1 x 50ml
50 x 1ml

## Shipping and storage

On arrival the kit should be stored at -20°C. Avoid prolonged exposure to light. If stored correctly the kit will retain full activity for 12 months. The kit can be stored at 4°C for 1 month. The kit can go through 30 freeze/thaw cycles with no loss of activity.

### Limitations of product use

The product may be used only for in vitro research purposes.

## Technical support

For technical support and troubleshooting please email technical@pcrbio.com the following information:

Amplicon size Reaction setup Cycling conditions Screen grabs of amplification traces and melting profile



#### Important considerations

Instrument compatibility: Different real-time PCR instruments require different levels of ROX passive reference. Generally, modern instruments do not require passive reference but include the option to use it for normalisation. Please check our qPCRBIO Selection Table to determine which ROX concentration your instrument requires (http://www.pcrbio.com/realtime-pcr.html).

Primer design: For efficient amplification under fast cycling conditions we recommend amplicon lengths between 80bp and 200bp. With all manufacturers master mixes the shorter the amplicon length the faster the reaction can be cycled. Amplicon lengths should not exceed 400bp. Primers should have a predicted melting temperature of around 60°C, using default Primer 3 settings (http://frodo.wi.mit.edu/primer3/). For TaqMan® probes choose probe close to 5' primer, avoid terminal guanosine residues.

Template amount: For genomic DNA,  $1\mu g$  or less is recommended. For cDNA, 100ng or less is recommended. However, users are encouraged to attempt a dilution series for new template/primer pairs to ensure that the PCR is efficient at that template concentration.

#### **Reaction setup**

- 1. Before starting, briefly vortex 2x qPCRBIO Probe Mix.
- 2. Prepare a master mix based on following table:

Reagent	20µl reaction	Final concentration	Notes	
2x qPCRBIO Probe Mix	10µl	lx		
Forward primer (10µM)	0.8µl	400nM	See above for optimal	
Reverse primer (10µM)	0.8µl	400nM	primer design	
Probe (10µM)	0.4µl	200nM		
Template DNA	<100ng cDNA, <1µg genomic	variable	See above for template considerations	
PCR grade dH <sub>2</sub> O	Up to 20µl final volume			

3. Program the instrument using following conditions, acquiring data on the appropriate channel:

Cycles	Temperature	Time	Notes
1	95°C	2min	Polymerase activation, 2 minutes for cDNA and 3 minutes for genomic
40	95°C 60°C to 65°C	5 seconds 20-30 seconds	Denaturation Anneal/Extension, do not exceed 30 seconds, do not use temperatures below 60°C
Melt analysis	It analysis Refer to instrument instructions		Optional melt profile analysis, available for hybridisation probes only