2.0X Taq RED Master Mix Kit (1.5mM $MgCl_2$) Cat #: 42-138B

Contents: 1000 Reactions

Storage: -20°C.
Reagent for *in vitro* laboratory use only

General Description

Apex Taq RED DNA Polymerase Master Mix from Genesee Scientific is a ready-to-use 2.0X reaction mix. Simply add primers, template, and water to successfully carry out primer extensions and other molecular biology applications. Apex Taq DNA Polymerase, the NH_4^+ buffer system, dNTPs and magnesium chloride are present in the Taq RED DNA Polymerase Master Mix. Each reaction requires 25 µl of the 2.0X reaction mix. Simply add primers, template and water to a total reaction volume of 50 µl.

Taq RED DNA Polymerase Master Mix offers several advantages. Set up time is significantly reduced. There is no need to buy and use separate loading dyes to load reaction products onto agarose gels for electrophoresis and subsequent visualization. The chance of contaminating component stocks is eliminated. Reduction of reagent handling steps leads to better reproducibility. Standard tests can be set up with the confidence that results will be consistent every time.

Composition of the 2.0X Taq RED Master Mix (1.5 mM $MgCl_2$ final concentration)

- Tris-HCl pH 8.5, $(NH_4)_2SO_4$, 3 mM $MgCl_2$, 0.2 % Tween® 20
- 0.4 mM of each dNTP
- Apex Taq DNA polymerase
- Inert red dye and stabilizer

Storage and Stability

Long term storage at -20 °C. Product expiry at -20 °C is stated on the label.
Option: Store at +4 °C for up to 6 months.

Quality Control

Taq DNA Polymerase is tested for contaminating activities, with no traces of endonuclease activity, nicking activity or exonuclease activity.

Protocol

This protocol serves as a guideline for primer extensions. Optimal reaction conditions such as incubation times, temperatures, and amount of template DNA may vary and must be individually determined.

Notes:

- Set up reaction mixtures in an area separate from that used for DNA preparation or product analysis.
- The table below shows the reaction set up for a final volume of 50 µL.
- Keep all components on ice.
- **Important:** Mix the solutions completely before use to avoid localized concentrations of salts.

1. Set up each reaction as follows:

Component	Vol./Reaction	Final Conc.
2.0X Taq RED Master Mix	25 µL	1X
Primer A	Variable	0.1–1.0 µM
Primer B	Variable	0.1–1.0 µM
Nuclease-Free Water	Variable	- - - -
Template DNA	Variable	Variable
TOTAL volume	50 µL	- - - -

2. Mix gently by pipetting the solution up and down a few times.
3. Program the thermal cycler according to the manufacturer's instructions.

For maximum yield and specificity, temperatures and cycling times should be optimized for each new template target or primer pair.

Table 3. Three-step PCR program

Cycles	Duration of cycle	Temperature
1	2-5 minutes	95 °C
25-35	20 – 30 seconds[a] 20-40 seconds[b] 30 seconds[c]	95 °C 55 – 60 °C 72 °C
1	5 minutes[d]	72 °C

[a.] Denaturation step: This step is the first regular cycling event and consists of heating the reaction to 95 °C for 20 – 30 seconds. It causes melting of the DNA template by disrupting the hydrogen

bonds between complementary bases, yielding single-stranded DNA molecules.

b. Annealing step: The reaction temperature is lowered to 50 – 65 °C for 20 – 40 seconds allowing annealing of the primers to the single-stranded DNA template. Typically, the annealing temperature is about 3 – 5 °C below the T_m (melting temperature) of the primers used.

c. Extension/elongation step: Taq polymerase has its optimum activity temperature at 72 °C. At this step the DNA polymerase synthesizes a new DNA strand complementary to the DNA template strand. The extension time depends on the length of the DNA fragment to be amplified. As a rule of thumb, at its optimum temperature the DNA polymerase will polymerize a thousand bases per minute.

d. Final elongation: This single step is occasionally performed at a temperature of 72 °C for 5 minutes after the last PCR cycle to ensure that any remaining single-stranded DNA is fully extended.

Related Products

Taq Polymerase kits (500 units)	Cat#
With 10X Standard and Ammonium Reaction Buffer	42-800B1
With 10X Combination Buffer	42-800B3
Glycerol Free	42-800B4

Hot Start DNA Polymerase (500 units)	Cat#
With 10X Ammonium and Combination Reaction Buffer	42-106

All polymerases are also available in kits, Mg^{2+} free buffers and 50 mM $MgCl_2$.

Master Mixes (500 reactions)	Cat#
2X Taq RED Master Mix, 1.5 mM $MgCl_2$	42-138
2X Taq Master Mix, Clear, 1.5 mM $MgCl_2$	42-134
2X Hot Start Master Mix Buffer I, 1.5 mM $MgCl_2$	42-198
2X Hot Start Master Mix Buffer I Blue, 1.5 mM $MgCl_2$	42-144

The shown Hot Start master mixes are ammonium based. Also available with balanced ammonium and potassium based buffers.

Real-time PCR (400 reactions)	Cat#
qPCR 2X Master Mix for Probe, without ROX™	42-116P
qPCR 2X Master Mix for Probe, low ROX™	42-118P
qPCR 2X Master Mix for Probe, high ROX™	42-120P
qPCR 2X GREEN Master Mix, without ROX™	42-116PG
qPCR 2X GREEN Master Mix, low ROX™	42-118PG
qPCR 2X GREEN Master Mix, high ROX™	42-120PG

DNA Ladders	Cat#
Apex 100 bp-Low DNA Ladder, 250 applications	19-109
Apex 1 kb DNA Ladder, 333 applications	19-115
Apex 200 bp DNA Ladder, 200 applications	19-111
ECON Mini DNA Ladder 100-500 bp, 100 applications	19-130
ECON Low DNA Ladder 100-1000 bp, 100 applications	19-131
ECON PCR Ladder 100-3000 bp, 100 applications	19-132

Ultrapure dNTPs	Cat#
dNTP set, 100 mM each: 250 µl of each dA, dC, dG and dT	42-410
dNTP Set, 100 mM each: 1 ml of each dA, dC, dG and dT	42-403
dNTP Mix 40 mM (1 x 500 µl): 10 mM each dA, dC, dG, dT	42-411
dNTP Mix 100 mM (2 x 1 ml): 25 mM each dA, dC, dG, dT	42-405
dNTP Mix 10 mM (10 x 1 ml): 2.5 mM each dA, dC, dG, dT	42-406

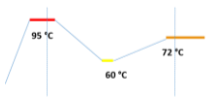
Other concentrations and Single dNTPs are available

Accessory reagents	Cat#
50 mM $MgCl_2$, 3 × 1.5 ml	42-303
Nuclease-Free Water, PCR Grade, 6 x 5 ml	42-710

Inspiration for Fast PCR

Fast PCR protocol:
Apex Taq DNA Polymerases and Taq DNA master mixes

90 minutes



Standard PCR protocol

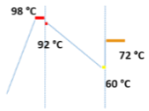
PCR program for 3-step Standard PCR

Cycler step	Temp.	Duration	Cycles
Initial heating	95 °C	3 min.	1
Denaturation	95 °C	30 sec.	
Annealing*	60 °C	30 sec.	30
Extension	72 °C	30 sec.	
Final extension	72 °C	5 min.	1

** the annealing temperature depends on the primer set.*

SAVE 1 HOUR - just change PCR cycler settings!

31 minutes

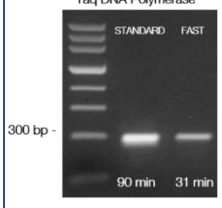


2-step Fast PCR protocol

PCR program for 2-step Fast PCR

Cycler step	Temp.	Duration	Cycles
Initial heating	98 °C	40 sec.	1
Denaturation	92 °C	2 sec.	
Extension*	60 °C	2 sec.	30
Final extension	72 °C	5 min.	1

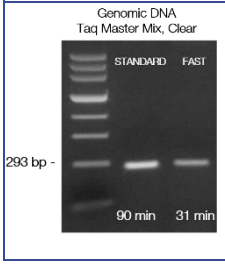
** the extension temperature depends on the primer set.*
For fast PCR choose highest possible T_m.



Amplification of λ DNA - Taq DNA Polymerase

Reaction mix*	
Ammonium buf.	1x
dNTP mix	0,2 mM each
$MgCl_2$	1,5 mM
Primers	0,2 µM
λ DNA	1 ng
Taq DNA pol.	0.5 – 1U

** H_2O up to a total volume of 25 µl*



Amplification of gDNA - 2x Taq Master Mix, Clear

Reaction mix*	
Taq MM Clear	1x
Primers	0,2 µM
gDNA	20 ng

** H_2O up to a total volume of 25 µl*